

Quantitative Evaluation of Mean Vascular Density using CD34 as Immunohistochemical Marker in Variants of Ameloblastoma

Hafiza Shahzadi Maryam¹, Alamgir², Shamsul Hadi³ and Sehrish⁴,

^{1,3}Department of Pathology, Peshawar Medical College.² Department of Oral and Maxillofacial Surgery, Ayub Teaching Hospital Abbottabad.⁴ Type D hospital Baffa Mansehra KP

ABSTRACT

Introduction: CD34 (Q-BEND 10) is a pan endothelial marker, monomeric glycoprotein and the cell surface trans membrane which is found in blood vessels of normal and neoplastic endothelial cells and use as a vascular marker for quantitative evaluation of angiogenesis. We determined the mean vascular density using CD34 as immunohistochemical marker in variants of Ameloblastoma.

Material and Method: This is a descriptive cross-sectional study of 6 months duration from June 2018-dec 2018. Sample was collected using consecutive sampling technique (non-probability sampling) and 30 Formalin fixed paraffin embedded (FFPE) blocks of Ameloblastoma were collected. The mean vascular density (MVD) in ameloblastoma was evaluated using method described by Nielsen and McNagny. Three areas with the highest amount of vascularization (known as the hot spot) were selected under low magnification ($\times 10$) and micro vessels were counted in each specimen at $\times 40$ magnification. The mean in three selected regions was considered as the MVD.

Results: The study consisted of already diagnosed 30 cases of ameloblastoma. The age range was between 20 - 40 years followed by 50 to 60 years with mean age 36.13 ± 16.0 . CD34 shows positive expression for all 30 cases. Among them 14 (46.7%) were males and 16 (53.3%) were females. The most common site of tumor was mandible in 83.3% cases followed by maxilla in 16.7%. The most common type was follicular ameloblastoma comprising of 16 (53.3%) cases, followed by plexiform 8 (26.7%), unicystic 5 (16.7%) while only 1 (3.3%) was of mixed type. The MVD range from 9.33 - 26.3. MVD of all cases was found to be 19.12 ± 7.8 .

Conclusion: It was concluded from this study that, CD34 can be used as a reliable endothelial marker to access MVD in different types of ameloblastoma.

Key Words: Ameloblastoma, Follicular ameloblastoma, Angiogenesis, CD34

Introduction

Ameloblastoma is a benign epithelial odontogenic tumor that morphologically resembles an enamel organ. According to the World Health Organization (1992) definition, ameloblastoma is a benign polymorphic tumor characterized by an increase in local invasive and odontogenic epithelium, the appearance of which is usually associated with follicular or plexiform and fibrous stroma.

The ratio of the mandible to the upper jaw varies from 80 to 20% and 99 to 1%. Most ameloblastomas of the mandible are located in the area of the molar ramus.¹

Annually, about 0.5% cases of ameloblastoma are diagnosed per million people worldwide² and odontogenic tumors are more common in Africa and China.³ Ameloblastoma is the second most common odontogenic tumor in the Western Hemisphere after squamous cell carcinoma. Therefore, African Americans are five times more likely than Caucasians to develop ameloblastoma than Caucasians with any other.⁴ Ameloblastomas account for 47.4 percent of all odontogenic tumors in Nigeria, according to the National Cancer Registry⁵. According to an Iranian study, ameloblastomas account for 42.5 percent of all odontogenic tumors⁶. Paediatric cases accounts for approximately 10 percent to 15 percent of all ameloblastoma cases in India, while in Africa and Asia, this percentage can reach as high as 25 percent of all cases⁷. A study conducted in Karachi, Pakistan, in

CORRESPONDENCE AUTHOR

Hafiza Shahzadi Maryam

Department of Pathology

Peshawar Medical College

2014 discovered that ameloblastoma accounted for 24.8 percent of all odontogenic tumors⁸.

The most commonly used classification system classifies ameloblastoma into the following categories based on clinical, radiological, and morphological criteria⁹.

- Solid/multicystic.
- Unicystic.
- Desmoplastic.
- Peripheral.
- Malignant.

The most common type of ameloblastoma is solid/multiple cystic and traditional, accounting for approximately 91% of all ameloblastoma cases. It grows slowly and takes benign path. Ameloblastoma is divided into two separate histological pattern: follicular and plexiform. Odontogenic cells of the follicular epithelium are grouped in islands, and plexiform epithelial cells are grouped in a continuous anastomosis row. Ameloblastoma does not often show all histological types.

The follicular type shows proliferating epithelial odontogenic cells organized in islands, while the plexiform type shows epithelial cells arranged in continuous anastomosing row. It is not rare for an ameloblastoma to exhibit all histological trends. In addition to these two histological forms, cystic, granular, acanthomatous, spindle cell, basal cell, clear cell and other microscopic subtypes have been identified. Unicystic ameloblastoma is the second most prevalent ameloblastoma and accounts for approximately 5–15 percent of all cases¹⁰. Unicystic Ameloblastoma have a relatively benign biologic behavior and better response to conservative treatment. Predominantly it is located in mandible, radiologically unilocular, and histologically it is characterized as a cystic lesion lined by an ameloblastomatous epithelial lining¹¹ and the more recent recommendations recognize two major histopathological forms of luminal and mural unicystic ameloblastoma¹². The luminal version reveals a cystic pattern lined with an ameloblastomatous epithelium protruding into the lumen as plexiform proliferations that appear like an intraluminal subtype. The wall variant reveals either the follicular or the plexiform configuration of the epithelial ameloblastomatous cells within the cyst wall. It is not rare for both variants to be found in the same ameloblastoma lesion.

Angiogenesis is triggered by a cocktail of growth factors and pro-angiogenic cytokines that modulates

another group of molecules blocking neovascularization. It is strictly regulated by balancing these pro-angiogenic growth factors and cytokines¹³.

Although Angiogenesis cannot be directly quantified, microvasculature can be determined by measuring MVD. The procedure involves immunohistochemical staining (IHC) of capillary endothelial cells with monoclonal antibodies¹⁴. Thus, MVD is useful for predicting metastasis and tumor recurrence, and angiogenesis which is critical for tumor growth, differentiation, development, and progression^{15,16}.

The MVD differentiation group (CD) is evaluated microscopically using various markers (monoclonal antibodies) such as “antibodies to CD105, CD34 and vascular endothelial growth factor (VEGF) and beta fibroblast growth factor¹⁷.”

CD34 is found in endothelial cells of normal and malignant blood vessels. It is used as a specific vascular marker to detect angiogenesis in various lesions, depending on its availability and ease of use¹⁸⁻²⁰.

Materials and Methods

This is descriptive cross-sectional study of six months duration from June 2018 to Dec 2018. The study consisted of already diagnosed formalin fixed paraffin embedded tissue (FFPE) sections of 30 cases of the ameloblastoma of different age and gender group. Formalin Fixed Paraffin Embedded (FFPE) representative tissue, blocks with the adequate tissue were included. FFPE with processing artifacts, blocks of true cut biopsy and non-representative tissues were excluded.

H&E slides of available blocks were reviewed by the pathologist. Five slides from representative blocks, 4 to 5 micron thin sections were made. Two of them were stained with H&E and two kept for IHC and one as reserve. Additional slides of the tonsil tissues was mounted for positive control.

IHC were carried out with the use of with the use of Mouse Anti-Human CD 34 Monoclonal Antibody (Clone: Q Bend 10, Product code: M7165 A/S, Glostrup, and DAKO, Denmark).”

The IHC staining of blood vessel in ameloblastoma was assessed by using technique described by (Nielsen & McNagny, 2008). The stained sections were first inspected at a low magnification¹⁰ in order to identify the locations with the strongest CD34 staining. Following that, blood vessel counting was carried out with a magnification of 40x. A cluster of endothelial cells with a lumen is termed a blood vessel those

endothelial cells are brown in color and positive for CD34. Three regions were chosen because they had the greatest amount of vascularization (known as hot spot). The mean of three selected areas was considered to be the MVD.

The objects which originate from one blood vessel were counted if it is completely separated from it. Blood vessels with muscular walls were eliminated. This was done by two pathologists blindly. In case of any disagreement opinion from third pathologist was taken.

After approval from the institutional Ethical committee of Peshawar Medical college and formal permission from Lab Administration was obtained for collecting blocks from the laboratories.

The data was analysed using the Statistical Package for Social Sciences (SPSS) version 22. Independent t-test was used to compare MVDs. Probability value ($p \leq 0.05$) was considered statistically significant.

Results

Majority of patients diagnosed with ameloblastoma were between the ages of 20 and 40 years, followed by those between 50 and 60 years, with a mean age of 36.13 ± 16.0 years.

Among these 30 cases 14 (46.7%) were males, 16 (53.3%) were females (Figure 1). Male to female ratio was 1:1.1. The most common site of the tumor was mandible in 25 (83.3%) cases while maxilla in only 5 (16.7%) cases.

The age range is between 12 and 80 years, with 36.13 ± 16.0 .

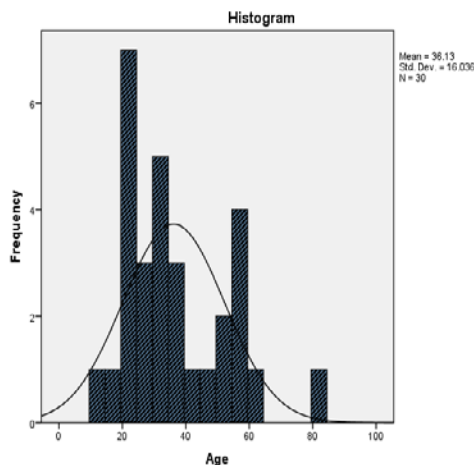


Figure 1 A histogram showing the mean age of the cases (n=30)

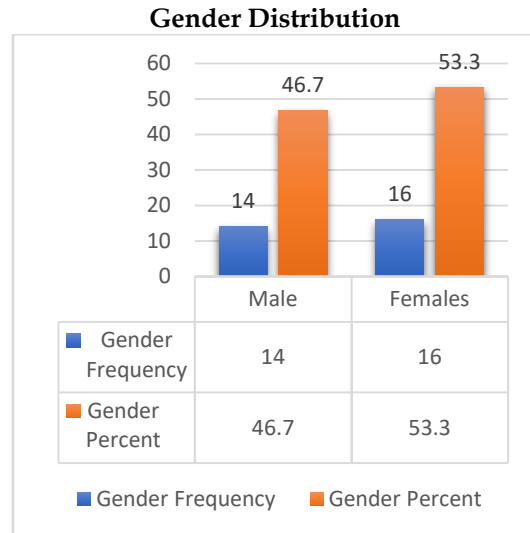


Figure 2 Gender distribution of Ameloblastoma cases (n=30)

Site of the Tumor:

The site of the tumor was mandible in 25 (83.3%) cases while maxilla in 5 (16.7%) cases

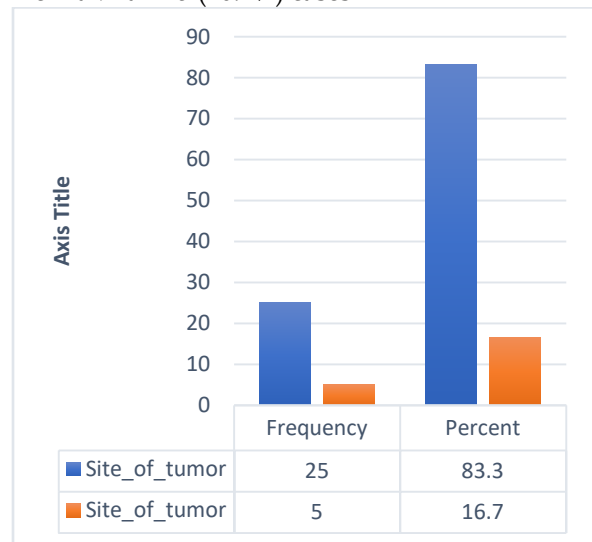


Figure 3 Frequency of the site of the tumor in ameloblastoma cases (n=30)

Among the 30 samples included in this study, 16 (53.3%) were follicular, 8 (26.7%) were plexiform, 5 (16.7%) were unicystic, while only 1 (3.3%) was of mixed type.

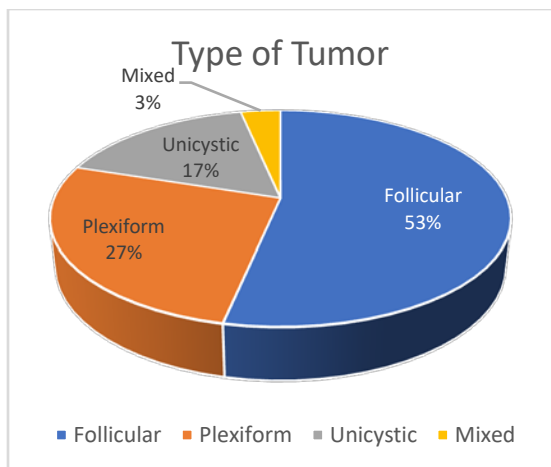


Figure 4. Frequency of the types of ameloblastoma (n=30)

The MVD in ameloblastoma ranged from 52 to 9.33 with a mean of 19.1 ± 7.8 .

Table-1: Mean Vascular Density:

The MVD in ameloblastoma ranged from 52 to 9.33 with a mean of 19.12 ± 7.8 .

S.NO	Vascular Density 1	Vascular Density 2	Vascular Density 3	MVD
1	26	24	20	23.33
2	36	16	27	26.33
3	15	11	7	11
4	21	25	16	20.66
5	19	12	13	14.66
6	13	15	11	13
7	52	46	58	52
8	14	17	12	14.33
9	26	18	17	20.33
10	17	15	19	17
11	18	26	22	22
12	20	23	18	20.33
13	13	12	19	14.66
14	20	31	33	28
15	21	16	22	19.66
16	16	11	18	15
17	26	33	19	26
18	14	24	13	17
19	22	38	18	26
20	11	9	8	9.33
21	16	25	18	19.66
22	16	13	11	13.33
23	17	15	24	18.66
24	16	13	14	14.33
25	15	10	16	13.66
26	18	18	13	16.33
27	10	13	9	10.66
28	12	17	18	15.66
29	20	20	20	20
30	19	25	18	20.66

Table 2: Relationship of MVD to ameloblastoma variants

Variants	No. of cases	Minimum MVD	Maximum MVD	Means \pm standard deviations
Follicular	16	13	52	20.89 ± 4.42
Plexiform	8	11	22	15.74 ± 2.53
Unicycstic	5	9.33	26.33	18.59 ± 5.38
Mixed	1	16	25	19.66 ± 4.36

Discussion

The result of the present study, shows staining positive for CD34 as in internationally published studies²¹⁻²² and a study published in Iran²³.

In the present study the common age group for occurrence of the ameloblastoma was second and third decade of life, with the mean age 36.13 ± 16.0 years. The results were similar to other reported studies^{24,25}, Santos et al, 2014 in their review of 112 cases of ameloblastoma reported a mean age of 35.1 years.

Male to female ratio of the disease is in correlation with study from Iran²³. The ratio of male to female was slightly higher than our study performed in Nigeria i.e 1.2: 1,²⁴ while in another study reported from Bangladesh male to female ratio was higher i.e 1.4:1²⁹.

In our study mandible was the commonest site of ameloblastoma (83.3%) followed by maxilla (16.7%) cases. The results are comparable with an international study. A study in Libya conducted showed comparable results with our study i.e Almost 92,8% of ameloblastomas were located in the mandible, with a very high mandible to maxilla ratio 13:1²⁶.

In current study follicular ameloblastoma was the most common histological variant followed by plexiform. These findings are consistent with the internationally published study in which follicular ameloblastoma was the common histological type (64.9%), plexiform (13.0%), desmoplastic ameloblastoma accounting for (5.2%), acanthomatous ameloblastoma shows squamous metaplasia accounted for (3.9%), and unicycstic ameloblastoma was diagnosed in (1.3%) cases²⁷.

Like other tumors ameloblastoma in our study showed a classic blood vessel-dependent pathology. The MVD of our cases was 19.12 ± 7.8 which was comparable with study done by Chen who found it to be 20.63 ± 7.30 ²⁸.

Among multicystic ameloblastomas follicular showed higher MVD as compared to plexiform, Unicycstic showed lesser MVD as compared to multicystic ameloblastoma. These findings were consistent with

the findings of the majority of studies, namely that the more aggressive the tumor that emerges from a certain tissue, the higher the MVD value of that tissue. Ameloblastomas possessed a dense vascular network, and the number of microvascular complications (MVDs) increased with recurrence and malignant transformation²⁹. In most odontogenic lesions, the distribution of blood vessels throughout all zones of the disease is not consistently the same. When comparing intratumoral MVD of multicystic ameloblastomas to peritumoral MVD, the intratumoral MVD exhibits the highest angiogenesis rate. An increase in MVD in the intratumoral region implies that odontogenic epithelial cells are responsible for angiogenesis in that region. It seems that the accumulation of blood vessels close by odontogenic epithelium for the provision of nutritional substances and oxygen is necessary for essential growth of odontogenic cysts and tumor²².

Angiogenesis has a prognostic significance and MVD is a useful marker to identify patients with ameloblastoma and aggressive ameloblastoma.

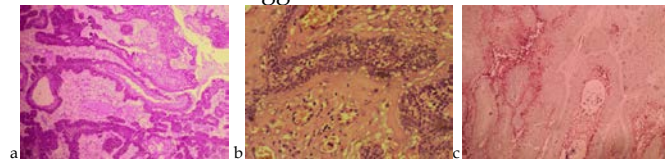


Figure 5. Follicular ameloblastoma a. (H & E 10X) b. (H & E 40x) arrow shows the vessel. c. Microvessels positive for immunohistochemical staining of CD 34 antigen for follicular ameloblastoma (10X)

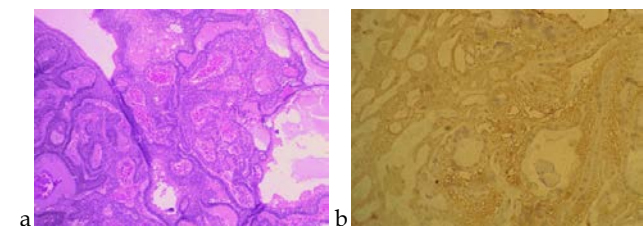


Figure 6 Plexiform ameloblastoma (H & E 10 X) b. Microvessels positive for immunohistochemical staining of CD 34 antigen for plexiform ameloblastoma (10X).

Conclusion

The study concluded that CD34 is a useful marker to assess angiogenesis and MVD. Follicular ameloblastoma showed a higher MVD as compared to other pointing at an aggressive behavior and close follow up for recurrence.

Research limitations

1. Small sample size
2. Financial constraint

Conflict of interest: None

Grant Support & Financial Disclosures: None

References

1. Giraddi GB, Bimleshwar SC, Singh C, Garg V, Anusha JS. Ameloblastoma—series of 7 treated cases—and review of literature. *Arch Oral Sci Res.* 2011; 1:152-5.
2. Brown and Betz. Ameloblastoma: A Review of Recent Molecular Pathogenetic Discoveries. *Biomarkers in Cancer* 2015;7(S2) 19-24 doi:10.4137/BIC.S29329.
3. Bansal S, Desai RS, Shirsat P, Prasad P, Karjodkar F, Andrade N. The occurrence and pattern of ameloblastoma in children and adolescents: an Indian institutional study of 41 years and review of the literature. *International journal of oral and maxillofacial surgery.* 2015 Jun 1; 44(6):725-31..
4. McClary AC, West RB, McClary AC, Pollack JR, Fischbein NJ, Holsinger CF, Sunwoo J, Colevas AD, Sirjani D. Ameloblastoma: a clinical review and trends in management. *European Archives of Oto-Rhino-Laryngology.* 2016 Jul; 273(7):1649-61.
5. Anyanechi CE, Saheeb BD. A review of 156 odontogenic tumours in Calabar, Nigeria. *Ghana medical journal.* 2014 Sep 18; 48(3):163-7.
6. Saghravanian N, Jafarzadeh H, Bashardoost N, Pahlavan N, Shirinbak I. Odontogenic tumors in an Iranian population: a 30-year evaluation. *Journal of oral science.* 2010; 52(3):391-6.
7. Bansal S, Desai RS, Shirsat P, Prasad P, Karjodkar F, Andrade N. The occurrence and pattern of ameloblastoma in children and adolescents: an Indian institutional study of 41 years and review of the literature. *International journal of oral and maxillofacial surgery.* 2015 Jun 1; 44(6):725-31.
8. Akram S, Anwar M, Shakir MM. Prevalence of odontogenic cysts and tumors in Karachi, Pakistan. *Journal of the Dow University of Health Sciences (JDUHS).* 2013 Apr 28;7(1):20-4.
9. Rastogi, S., Nijhawan, S., Modi, M., Kumar, A., Aslam, N., Lstheef, F. J. J. o. C., & Research, d. (2010). Radiolucent-radiopaque lesion in the mandible—a nobel diagnostic approach. 4, 2300-2307.
10. Dhanuthai K, Chantarangsu S, Rojanawatsirivej S, Phattararatip E, Darling M, Jackson-Boeters L, Said-Al-Naief N, Shin HI, An CH, Hong NT, An PH. Ameloblastoma: a multicentric study. *Oral surgery, oral medicine, oral pathology and oral radiology.* 2012 Jun 1; 113(6):782-8.
11. Siriwardena BS, Tennakoon TM, Hunter KD, Tilakaratne WM. Unicystic ameloblastoma: Analysis of

370 cases in a single center in Sri Lanka. *Journal of Oral Pathology & Medicine*. 2018 Aug; 47(7):706-9.

12. Wright JM, Vered M. Update from the 4th edition of the World Health Organization classification of head and neck tumours: odontogenic and maxillofacial bone tumors. *Head and neck pathology*. 2017 Mar; 11(1):68-77.
13. Naumov GN, Bender E, Zurakowski D, Kang SY, Sampson D, Flynn E, Watnick RS, Straume O, Akslen LA, Folkman J, Almog N. A model of human tumor dormancy: an angiogenic switch from the nonangiogenic phenotype. *Journal of the National Cancer Institute*. 2006 Mar 1; 98(5):316-25.
14. Pandiar D, Shameena PM. Immunohistochemical expression of CD34 and basic fibroblast growth factor (bFGF) in oral submucous fibrosis. *Journal of Oral and Maxillofacial Pathology: JOMFP*. 2014 May; 18(2):155.
15. Folkman J. Fundamental concepts of the angiogenic process. *Current molecular medicine*. 2003 Nov 1; 3(7):643-51.
16. [16] Weidner N, Semple JP, Welch WR, Folkman J. Tumor angiogenesis and metastasis—correlation in invasive breast carcinoma. *New England Journal of Medicine*. 1991 Jan 3;324(1):1-8.
17. Kademani D, Lewis JT, Lamb DH, Rallis DJ, Harrington JR. Angiogenesis and CD34 expression as a predictor of recurrence in oral squamous cell carcinoma. *Journal of oral and maxillofacial surgery*. 2009 Sep 1; 67(9):1800-5.
18. Ancuța CO, Ancuța E, Zugun-Eloae F, Carasevici E. Neoangiogenesis in cervical cancer: focus on CD34 assessment. *Rom J Morphol Embryol*. 2010 Jan 1;51(2):289-94.
19. Chalooob MK, Ali HH, Qasim BJ, Mohammed AS. Immunohistochemical expression of Ki-67, PCNA and CD34 in astrocytomas: a clinicopathological study. *Oman medical journal*. 2012 Sep;27(5):368.
20. Segatelli V, de Oliveira EC, Boin IF, Ataide EC, Escanhoela CA. Evaluation and comparison of microvessel density using the markers CD34 and CD105 in regenerative nodules, dysplastic nodules and hepatocellular carcinoma. *Hepatology international*. 2014 Apr;8(2):260-5.
21. Nielsen JS, McNagny KM. Novel functions of the CD34 family. *Journal of cell science*. 2008 Nov 15;121(22):3683-92.
22. Seifi S, Shafaie S, Ghadiri S. Microvessel density in follicular cysts, keratocystic odontogenic tumours and ameloblastomas. *Asian Pac J Cancer Prev*. 2011 Jan 1;12(2):351-6.
23. Jamshidi S, Zargaran M, Baghaei F, Shojaei S, Zare Mahmoodabadi R, Dehghan A, et al. An immunohistochemical survey to evaluate the expression of CD105 and CD34 in ameloblastoma and odontogenic keratocyst. *J Dent (Shiraz)* 2014;15:192-8.
24. Agbaje JO, Adisa AO, Petrova MI, Olusanya AA, Osayomi T, Effiom OA, Soyele OO, Omitola OG, Olawuyi AB, Okiti RO, Saiki TE. Biological profile of ameloblastoma and its location in the jaw in 1246 Nigerians. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2018 Nov 1;126(5):424-31.
25. Seifi S, Feizi F, Khafri T, Aram M. Histomorphometric comparative study of blood vessels and their pattern in follicular cyst, odontogenic keratocyst, and ameloblastoma. *Journal of Craniofacial Surgery*. 2013 Mar 1;24(2):439-45.
26. Rahman SB, Sadat SA, Haider IA, Ahmed M. Analysis of histological variants of ameloblastomas of jaws in relation to their clinical presentations. *Journal of Bangladesh College of Physicians and Surgeons*. 2017 Jul 29;35(2):61-7.
27. Adebisi KE, Ugboko VI, Omoniyi-Esan GO, Ndukwe KC, Oginni FO. Clinicopathological analysis of histological variants of ameloblastoma in a suburban Nigerian population. *Head & Face Medicine*. 2006 Dec; 2(1):1-8.
28. Feng CW, Wang LD, Jiao LH, Liu B, Zheng S, Xie XJ. Expression of p53, inducible nitric oxide synthase and vascular endothelial growth factor in gastric precancerous and cancerous lesions: correlation with clinical features. *BMC cancer*. 2002 Dec;2(1):1-7.
29. Bossi P, Viale G, Lee AK, Alfano R, Coggi G, Bosari S. Angiogenesis in colorectal tumors: microvessel quantitation in adenomas and carcinomas with clinicopathological correlations. *Cancer research*. 1995 Nov 1; 55(21):5049-53.

HISTORY	
Date received:	18-8-2022
Date sent for review:	22-8-2022
Date received reviewers comments:	30-8-2022
Date received revised manuscript:	12-11-2022
Date accepted:	24-12-2022

CONTRIBUTION OF AUTHORS	
Author	Contribution
Hafiza Shahzadi Maryam	A,B,C
Alamgir	B
Shamsul Had	C
Sehrish	B

KEY FOR CONTRIBUTION OF AUTHORS:

- A. Conception/Study/Designing/Planning
- B. Active Participation in Active Methodology
- C. Interpretation/ Analysis and Discussion